

Fabrication of CNT microarray for biosensor applications

Nji Raden Poespawati^{a*}, Tomy Abuzairi^{a,b}, Mitsuru Okada^c, Retno Wigajatri Purnamaningsih^a, Masaaki Nagatsu^{b,c,d}

^aDepartment of Electrical Engineering, Universitas Indonesia, 16424, Depok, Indonesia ^bGraduate School of Science and Technology, Shizuoka University, 432-8561, Hamamatsu, Graduate School of Integrated Science and Technology, Shizuoka University, 432-8561, Hamamatsu, Japan ^dResearch Institute of Electronics, Shizuoka University, 432-8561, Hamamatsu, Japan

Abstract

Microarray technology has become one of the indispensable tools which can be used to identification of bio-molecules in cells, tissues, and disease, such as disease diagnosis, prediction, prevention, and drug discovery. The major advantages of the microarray biosensor are their ability of the simultaneous analysis of thousands parameters on a single platform and minimal sample consumption. Furthermore, carbon nanotubes (CNTs) with their outstanding properties are potential material for many applications including biosensors. To increase the fluorescence detection, the microarray will be established within the three-dimensional (3D) structure. The advantage of the 3D structure CNT for microarray platform is the enhancement of active surface area without sacrificing the size of the device. For instance, the external surface area of one CNT spot is roughly $\sim 3.14 \times 10^{-5}$ mm². As arrays of 100 mm² of CNT platform with 50 μ m spacing, it can contain as many as 1.82×10^8 spots that is high-density microarrays. This high-density microarray is powerful tools for the screening of pharmaceuticals, investigation of biomolecule interactions and patient diagnostics. In this paper, the CNT is fabricated in microarray configuration to realize biochip sensor design. The CNT microarray was fabricated by using electron beam lithography for patterning microarray on silicon substrate, RF sputtering for deposit catalyst, and thermal plasma chemical vapor deposition (CVD) for growing vertically aligned CNT.

Keywords

APPJ; plasma functionalization; CNT; 3D microarray; biosensor applications

^{*} Corresponding author. Tel.: +62-21-7270078; fax: +62 – 21- 7270077. *E-mail address:* <u>pupu@eng.ui.ac.id</u>



1. Introduction

Carbon nanotubes (CNTs) are nanomaterial carbon allotropes with a high surfacearea-to-weight ratio (~300 m²g⁻¹) (J. N. Wohlstadter et al., 2003; L. Dong et al., 1996) and excellent material properties such as electrical conductivities and mechanical strength (M. S. Dresselhaus & P. Avouris, 2001; P. M. Ajayan, 1999). CNTs are attracting much attention due to their outstanding properties, with promising applications in numerous areas such as field-effect transistor (S. J. Tans, et al., 1998), fuel cell (Wang, et al., 2004), lithium-ion battery (C. de las Casas & W. Li, 2012), transparent conductive film (Wu, et al., 2004), including biosensors (F. Lu et al., 2009).

Biosensor based on CNTs compose of three associated elements: (1) bioreceptor; (2) interface (functionalized CNT); and (3) transducer. Bioreceptor is a sensitive biological element (such as enzyme, DNA probe, or antibody) for recognizing the analyte (such as complementary DNA and antigen). To work as biosensor, the CNTs have to functionalize as interface layer because the CNTs structure is remarkably stable so that it is insoluble in most solvents. As a result, functionalization of CNT is necessary to allow a better processing of CNT toward the development of biosensors, and improve the solubility and the selectivity of biomolecules recognition (T. Abuzairi et al., 2015, 2016). The transducer converts the biological event to other measurable signals, such as currents, absorbance, mass or acoustic variables.

In this work, the CNT is fabricated in microarray configuration to realize biochip sensor design. The CNT microarray was fabricated by using electron beam lithography for patterning microarray on silicon substrate, RF sputtering for deposit catalyst, and thermal plasma chemical vapor deposition (CVD) for growing vertically aligned CNT (T. Matsuda, et al., 2009; T. Matsuda, et al., 2008).

2. Experimental

Initially, a silicon substrate 10×10 -mm² was cleaned by piranha solution (40 mL sulfuric acid and 10 mL hydrogen peroxide) for 3 minutes, and rinsed by a mixture of 47.5 mL distilled ionized (DI) water and 2.5 mL hydrofluoric acid (HF) for 1 minute. The Si substrate was then coated with hexamethyldisilazane (HMDS) as pre-resist coating and ZEP-520A as positive electron-beam photoresist by automatic spin coating Suss Delta 80 Gyrset. After that, the Si substrate was baked at 180°C for 2 minutes. The photoresist layer was patterned by electron beam lithography ELS-7700K with acceleration volatage 80 kV, beam current 1.5×10^{-9} A, and dose time 0.2 µs. To remove pattern photoresist, the substrate was developed by 0-Xylen at 23°C for 2 minutes and 2-propanol for 40 seconds. The pattern result was shown in **Fig. 1(a)**.

The substrate was then deposited by chromium (Cr) thin film as a diffusion barrier layer for 20 seconds and nickel (Ni) film as a catalyst for 60 seconds by a RF magnetron sputtering SVC-700RF in argon gas ambient as illustrated in **Fig.1(b)**. A lift-off process was conducted to remove the positive photoresist film by remover ZDMAC and left behind the array catalyst on the substrate after one day as depicted in **Fig. 1(c)**. Finally, the growth of CNT array was carried out in the plasma thermal CVD at a substrate temperature of 700 °C for 5 min pre-treatment with NH₃ (150 sccm) followed by 15 min



post-treatment with a mixture of $NH_3:C_2H_2$ (3:1) and an applied dc bias voltage of -550 V to the substrate stage.



Figure 1. Fabrication of carbon nanotube microarray. (a) Electron beam lithography. (b) RF sputtering. (c) Lift-off process.

3. Results and Discussion

The CNT were fabricated by using electron beam lithography for patterning microarray, RF sputtering for deposit nickel catalyst, and thermal plasma chemical vapor deposition for growing vertically aligned CNT on the silicon substrate (T. Matsuda et al., 2008). **Fig. 2** depicts an FE-SEM image of the CNT microarray with 50 μ m spacing and 5 μ m dot size, and magnified image of bundle structure of vertically aligned CNT in cross-section and top views. Each dot consisted of a bundle of multi-walled CNT having a typical length of ~2 μ m and diameter of about 100 nm. The vertically aligned CNT were fabricated in microarray form to realize the development of biosensor (Kricka, 2001), which has a comparable size of the microarrays biosensor fabricated by other methods, for example photolithography (Malainou, et al., 2012), physical masking (Lee et al., 2010), laser printing method (Duocastella, et al., 2010), micromachine (Bhatnagar, 2007), or microcantilivers processing (Belaubre et al., 2003).

Each CNT spot is comprised of a bundle of multiwall carbon nanotube having a typical length of about 2 μm and a diameter of about 100 nm. The primary advantage of



the 3D structure CNT for biosensor platform is the enhancement of active surface area without sacrificing the size of the device. For instance, the external surface area of one CNT spot is roughly $\sim 3.14 \times 10^{-5}$ mm², calculated from the diameter and length of the CNT (5 µm diameter and 2 µm length) (A. Peigney, et al., 2001). As arrays of 100 mm² of CNT platform with 50 µm spacing, it can contain as many as 1.82×10^8 spots that is high-density biochip microarrays. This high-density microarray is powerful tools for the screening of pharmaceuticals, investigation of biomolecule interactions and patient diagnostics (C. Wingren & C. A. Borrebaeck, 2007; J. Clausmeyer, et al., 2014).



Figure 2. FE-SEM images of the CNT biochip sensor platform and a magnified image of one spot CNT is shown inset (T. Abuzairi et al., 2016).

Additionally, based on the calculation of the CNT surface area, the total surface area of one platform is estimated as ~5715 mm². Therefore, the 3D structure of CNT will enhance surface area as ~57 times compared with the 2D structure platform of 100 mm². From this point of view, CNT materials are great potential for miniaturization the biochip device, which may allow for the detection of currently undetectable disease markers (C. Wingren & C. A. Borrebaeck, 2007; J. W. Silzel, et al., 1998).

4. Conclusion

The vertically aligned CNT is successfully fabricated by using electron beam lithography for patterning microarray, RF sputtering for deposit nickel catalyst, and thermal plasma chemical vapor deposition (CVD) on the silicon substrate. The 3D structure of vertically aligned CNTs can enhance the active surface area without sacrificing the size of the device. Therefore, CNT materials are great potential for miniaturization the microarray biosensor, which may allow for the detection of currently undetectable disease markers.

Acknowledgements

This work has been supported by International Research Collaboration and Scientific Publication Grant (DIPA – 042.06.1.401516/2016) from Directorate General of Higher



Education (DGHE) Indonesia. The authors also acknowledge the support of Grant-in-Aid for Scientific Research (A) (No. 25246029) and 209 Scientific Research (B) (No. 26289016) from the Japan Society for the Promotion of Science.

References

- A. Peigney, C. Laurent, E. Flahaut, R. R. Bacsa, & A. Rousset. (2001). Specific surface area of carbon nanotubes and bundles of carbon nanotubes. *Carbon*, *39*(4), 507–514.
- Belaubre, P., Guirardel, M., Garcia, G., Pourciel, J. B., Leberre, V., Dagkessamanskaia, A., ... Bergaud, C. (2003). Fabrication of biological microarrays using microcantilevers. *Applied Physics Letters*, 82(18), 3122–3124.
- Bhatnagar, P. (2007). Multiplexed electrospray deposition for protein microarray with micromachined silicon device. *Applied Physics Letters*, *91*(1), 14102.
- C. de las Casas, & W. Li. (2012). A review of application of carbon nanotubes for lithium ion battery anode material. *A Review of Application of Carbon Nanotubes for Lithium Ion Battery Anode Material*, 28, 74–85.
- C. Wingren, & C. A. Borrebaeck. (2007). Progress in miniaturization of protein arrays a step closer to high-density nanoarrays. *Drug Discovery Today*, *12*(19), 813–819.
- Duocastella, M., Fernández-Pradas, J. M., Morenza, J. L., Zafra, D., & Serra, P. (2010). Novel laser printing technique for miniaturized biosensors preparation. *Sensors and Actuators B: Chemical*, 145(1), 596–600.
- F. Lu, L. Gu, M. J. Meziani, X. Wang, P. G. Luo, L. Veca, ... Y.-P. Sun. (2009). Advances in Bioapplications of Carbon Nanotubes. *Advances in Bioapplications of Carbon Nanotubes*, *21*(2), 139152.
- J. Clausmeyer, W. Schuhmann, & N. Plumeré. (2014). Electrochemical patterning as a tool for fabricating biomolecule microarrays. *TrAC Trends in Analytical Chemistry*, *58*, 23–30.
- J. N. Wohlstadter, J. L. Wilbur, G. B. Sigal, H. A. Biebuyck, M. A. Billadeau, L. Dong, ... S. J. Wohlstadter. (2003). Carbon nanotube-based biosensor. *Advanced Materials*, *15*(14), 1184–1187.
- J. W. Silzel, B. Cercek, C. Dodson, T. Tsay, & R. J. Obremski. (1998). Mass-sensing, multianalyte microarray immunoassay with imaging detection. *Clinical Chemistry*, 44(9), 2036–2043.
- Kricka, L. J. (2001). Microchips, microarrays, biochips and nanochips: personal laboratories for the 21st century. *Clinica Chimica Acta*, 307(1–2), 219–223.
- L. Dong, A. B. Fischer, M. Lu, M. T. Martin, D. Moy, & D. Simpson. (1996). Reversible and irreversible immobilization of enzymes on graphite fibrilsTM. *Journal of Molecular Recognition*, 9(5-6), 383–388.
- Lee, H.-U., Park, S.-Y., Kang, Y.-H., Jeong, S.-Y., Choi, S.-H., Jahng, K.-Y., & Cho, C.-R. (2010). Surface modification of and selective protein attachment to a flexible microarray pattern using atmospheric plasma with a reactive gas. *Acta Biomaterialia*, 6(2), 519–525.
- Malainou, A., Petrou, P. S., Kakabakos, S. E., Gogolides, E., & Tserepi, A. (2012). Creating highly dense and uniform protein and DNA microarrays through photolithography and plasma modification of glass substrates. *Biosensors and Bioelectronics*, *34*, 273–281.
- M. S. Dresselhaus, & P. Avouris. (2001). Introduction to carbon materials research. In *Carbon Nanotubes* (pp. 1–9). Springer.
- P. M. Ajayan. (1999). Nanotubes from Carbon. Nanotubes from Carbon, 99(7), 1787-1800.
- S. J. Tans, A. R. M. Verschueren, & C. Dekker. (1998). Room-temperature transistor based on a single carbon nanotube. *Room-Temperature Transistor Based on a Single Carbon Nanotube*, 393(6680), 49–52.
- T. Abuzairi, M. Okada, R. W. Purnamaningsih, N. R. Poespawati, F. Iwata, & M. Nagatsu. (2016). Maskless localized patterning of biomolecules on carbon nanotube microarray functionalized by ultrafine atmospheric pressure plasma jet using biotin-avidin system. *Applied Physics Letters*, 109, 23701.
- T. Abuzairi, M. Okada, Y. Mochizuki, N. R. Poespawati, R. W. Purnamaningsih, & M. Nagatsu. (2015). Maskless functionalization of a carbon nanotube dot array biosensor using an ultrafine atmospheric pressure plasma jet. *Carbon*, *89*, 208–216.



- T. Matsuda, J. Sato, T. Ishikawa, A. Ogino, & M. Nagatsu. (2009). Field emission characteristics of nano-sized dot array carbon nano-tube emitters fabricated by direct current plasma chemical vapor deposition. *Field Emission Characteristics of Nano-Sized Dot Array Carbon Nano-Tube Emitters Fabricated by Direct Current Plasma Chemical Vapor Deposition*, 18(2– 3), 548–552.
- T. Matsuda, M. Mesko, A. Ogino, & M. Nagatsu. (2008). Synthesis of vertically aligned carbon nanotubes on submicron-sized dot-catalyst array using plasma CVD method. *Diamond and Related Materials*, *17*(4–5), 772–775.
- Wang, C., Waje, M., Wang, X., Tang, J., & Haddon, R. (2004). Proton exchange membrane fuel cells with carbon nanotube based electrodes. *Proton Exchange Membrane Fuel Cells with Carbon Nanotube Based Electrodes*, 4(2), 345–348.
- Wu, Z., Chen, Z., Du, X., Logan, J., & Sippel, J. (2004). Transparent, conductive carbon nanotube films. *Transparent, Conductive Carbon Nanotube Films*, *305*(5688), 1273–1276.