

## [ Chiba Medical Society Award Review ]

## The role of transcription factor in the regulation of autoimmune diseases

## Akira Suto<sup>1,2)</sup>

<sup>1)</sup> Department of Allergy and Clinical Immunology, Graduate School of Medicine, Chiba 260-8670. <sup>2)</sup> Institute for Global Prominent Research, Chiba University, Chiba 260-8670.

(Received November 14, 2020, Accepted December 9, 2020, Published April 10, 2021.)

## Abstract

IL-21 is produced by Th17 cells and follicular helper T cells. It is an autocrine growth factor and plays critical roles in autoimmune diseases. In autoimmune mouse models, excessive production of IL-21 is associated with the development of lupus-like pathology in Sanroque mice, diabetes in NOD mice, autoimmune lung inflammation in Foxp3-mutant scurfy mice, arthritis in collagen-induced arthritis, and muscle inflammation in experimental autoimmune myositis. IL-21 production is induced by the transcription factor c-Maf following Stat3 activation on stimulation with IL-6. Additionally, Stat3 signaling induces the transcription factor Sox5 that along with c-Maf directly activates the promoter of RORyt, a master regulator of Th17 cells. Moreover, T cell-specific Sox5-deficient mice exhibit decreased Th17 cell differentiation and resistance to experimental autoimmune encephalomyelitis and delayedtype hypersensitivity. Another Sox family gene, *Sox12*, is expressed in regulatory T cells (Treg) in dextran sulfate sodium-induced colitic mice. T cell receptor-NFAT signaling induces Sox12 expression that further promotes Foxp3 expression in CD4<sup>+</sup> T cells. In vivo, Sox12 is involved in the development of peripherally induced Treg cells under inflammatory conditions in an adoptive transfer colitis model. This review highlights the crucial roles of transcription factors in the onset of autoimmune diseases and the differentiation of IL-21-producing CD4<sup>+</sup> T cells, Th17 cells, and Treg cells.

Key words: autoimmune disease, IL-21, c-Maf, Sox5, Sox12

### I. Introduction

Autoimmune diseases are caused by the breakdown of tolerance to self-antigens and are characterized by the activation of T cells, including pathogenic IL-

Phone: +81-43-226-2198. Fax: +81-43-226-2199 E-mail: suaki@faculty.chiba-u.jp 17-producing helper T cells (Th17 cells) and IL-21producing CD4<sup>+</sup> T cells. Th17 cells are eventually differentiated in the presence of IL-23 and IL-21, and further produce IL-17A, IL-17F, and IL-21. On the other hand, regulatory T (Treg) cells, defined by the expression of Foxp3, play a central role in protecting against excessive inflammatory responses caused by autoimmune diseases. This review highlights the crucial roles of transcription factors in the onset of autoimmune diseases and the transcriptional regulation of the

Address correspondence to Dr. Akira Suto.

Department of Allergy and Clinical Immunology, Graduate School of Medicine, Chiba University, 1-8-1 Inohana, Chiba 260-8670, Japan.

differentiation of Th17 cells, IL-21-producing  $CD4^+$  T cells, and Treg cells.

## II. Role of IL-21 in the onset of autoimmune diseases

IL-21 is a four-helix-bundle type I cytokine with significant homology to IL-2, IL-4, and IL-15[1]. *In vivo*, IL-21 has been shown to be involved in many kinds of autoimmune disease models. For example, lymphopenia induced IL-21-mediated homeostatic expansion has been shown to develop type I diabetes in NOD mice[2,3]. Excessive production of IL-21 is associated with high titers of auto-antibodies and development of lupus in *Sanroque* mice[4]. Furthermore, neutralization of IL-21 by IL-21-receptor (IL-21R) Fc chimera protein has been reported to ameliorate conditions in mouse models of lupus and rheumatoid arthritis[5,6].

Mutations in Foxp3 gene lead to development of IPEX (Immune dysregulation, polyendocrinopathy, enteropathy, X-linked) syndrome in humans[7]. Scurfy mice have a frame-shift mutation in Foxp3 gene. They completely lack Treg cells and suffer from autoimmune diseases, such as skin, lung, and liver inflammation, leading to death within 4 weeks of birth[8]. Activation of CD4<sup>+</sup> T cells caused by the lack of Treg cells plays a crucial role in the onset of autoimmune diseases in scurfy mice[9]. Among the subset of activated CD4<sup>+</sup> T cells, unique IL-21-producing c-Maf<sup>+</sup> CD4<sup>+</sup> T cells develop, which further induce short-lived effector CD8<sup>+</sup> T cells (SLEC) and accelerate multi-organ autoimmune inflammation in scurfy mice[10]. IL-21 receptor deficiency results in reduced multi-organ inflammation, prolonged survival, and decrease of SLEC in the lung in scurfy mice. Thus, IL-21-producing CD4<sup>+</sup> T cells are extensively involved in the onset of autoimmune lung inflammation in scurfy mice, presumably through the induction of SLEC. Further, Sharma et al. have shown that intramuscular injection of lymph node (LN) cells from scurfy mice into recombination activating gene (RAG)-deficient mice causes inflammatory myopathy (IM)-like pathological changes in uninjected muscles

[11,12]. This suggests the possible involvement of IL-21 in the pathogenesis of IM. Polymyositis (PM) and dermatomyositis (DM) are the major subgroups of idiopathic IM characterized by infiltration of inflammatory cells into the affected muscles[13]. Regarding the relationship between IL-21 and IM in humans, CXCR3<sup>-</sup> CXCR5<sup>+</sup> helper T cells, which produce IL-21, increase in peripheral blood in juvenile DM patients having skin rash and muscular weakness [14]. In addition, serum levels of IL-21 are increased in the subset of patients with IM as compared to those in healthy controls[15].

In experimental autoimmune myositis (EAM) IM mouse model, the severity of EAM is diminished in IL-21<sup>-/-</sup> mice[15]. GM-CSF production from  $\gamma\delta T$ cells, but not Th17 cell differentiation, is significantly reduced in EAM-induced IL-21<sup>-/-</sup> mice. Importantly, the neutralization of GM-CSF or deficiency of γδT cells significantly improves muscle weakness and muscle inflammation in EAM-induced mice. Major GM-CSF producers in EAM-induced mice muscles are  $V\gamma 4^+$  $V\delta4^+$  CD27<sup>-</sup> cells that are significantly decreased in EAM-induced IL-21<sup> $^{-/-}$ </sup> mice. Intriguingly, V $\gamma$ 4<sup>+</sup>  $V\delta4^+$  CD27<sup>-</sup> cells express high levels of CX3CR1. In addition, CX3CL1, a ligand of CX3CR1, is induced in the muscle upon EAM induction in WT mice but not in IL-21<sup>-/-</sup> mice. In summary, IL-21 facilitates autoimmune myositis through the accumulation of GM-CSF-producing  $V\gamma 4^+ V\delta 4^+$  cells in the muscle possibly via the CX3CR1-CX3CL1 pathway.

## III. Development of IL-21-producing $CD4^+$ T cells

Activated CD4<sup>+</sup> T cells differentiate into at least three distinct helper T cell subsets as defined by their patterns of cytokine production. Th1 cells produce IFN- $\gamma$  and lymphotoxin and protect against intracellular pathogenic infections. Th2 cells produce IL-4, IL-5, and IL-13 and are essential for defending against parasites. On the other hand, Th17 cells produce IL-17A and IL-17F and play a pathogenic role in a variety of autoimmune diseases. IL-21-producing CD4<sup>+</sup> T cells develop preferentially in Th17-polarizing condition (IL-6 + TGF- $\beta$  + anti-IL-4 antibody + anti-IFN- $\gamma$  antibody). Among these, IL-6 + anti-IL-4 antibody + anti-IFN- $\gamma$  antibody strongly induce the development of IL-21-producing CD4<sup>+</sup> T cells without inducing the production of IFN- $\gamma$ , IL-4, IL-5, IL-13, IL-17A, and IL-17F[16]. While TGF- $\beta$  inhibits IL-6-mediated differentiation of IL-21-producing CD4<sup>+</sup> T cells in a dose-dependent manner, IL-21 itself induces the differentiation of IL-21-producing CD4<sup>+</sup> T cells.

## IV. Transcriptional regulation of IL-21producing $CD4^+$ T cells

NFAT directly binds to the IL-21 promoter and activates its transcription in Th2 cells[17]. On the other hand, T-bet suppresses IL-21 transcription by inhibiting the binding of NFAT to the promoter in Th1 cells. IRF-4 binds to and activates IL-21 promoter; whereas, IRF-4-binding protein (IBP) inhibits IL-21 production by regulating the activity of IRF-4[18]. Stat3 (activated by IL-6 and/or IL-21) is required for IL-21 production in Th17 cells[19]. However, RORyt, the master regulator of Th17 cells, is not involved in the production of IL-21[19]. To determine the transcription factor that induces IL-21-producing CD4<sup>+</sup> T cells, DNA

microarray analysis of IL-6-stimulated CD4<sup>+</sup> T cells was performed. Expression of c-Maf, a basic leucine zipper protein, is significantly upregulated in IL-21producing CD4<sup>+</sup> T cells[20]. Overexpression of c-Maf strongly induces IL-21-producing CD4<sup>+</sup> T cells without IL-6 stimulation or autocrine effect of IL-21. c-Maf directly binds to and activates both IL-21 promoter and CNS-2 enhancer through Maf recognition elements (MAREs), TGCn<sub>6-8</sub>GCA (Fig. 1). On the other hand, TGF-β upregulates IL-6-induced c-Maf expression but inhibits c-Maf-induced IL-21 production in CD4<sup>+</sup> T cells. Additionally, Foxp3 binds to IL-21 promoter and CNS-2 enhancer and inhibits c-Maf-induced IL-21 production in CD4<sup>+</sup> T cells. In summary, c-Maf directly induces IL-21 production by activating IL-21 promoter and CNS-2 enhancer and TGF-B suppresses c-Mafmediated IL-21 production in CD4<sup>+</sup> T cells, presumably through Foxp3.

# V. Transcriptional regulation of Th17 cell differentiation

Th17 cells produce IL-17A and IL-17F and play a pathogenic role in many autoimmune diseases. Since ectopic expression of RORγt (encoded by *Rorc*) induces



Fig. 1 Roles of Sox5 and c-Maf in the production of IL-17A and IL-21. Under Th17-differentiating conditions, IL-6 induces Sox5 expression in CD4<sup>+</sup> T cells in a Stat3-dependent manner. Additionally, IL-6 induces c-Maf expression especially in the presence of TGF- $\beta$ . Sox5 and c-Maf co-induce IL-17A production via induction of ROR $\gamma$ t. c-Maf induces IL-21 by binding to the IL-21 promoter and CNS-2 enhancer. On the other hand, TGF- $\beta$  inhibits c-Maf-induced IL-21 production in CD4<sup>+</sup> T cells. TGF- $\beta$ -induced Foxp3 binds to IL-21 promoter and CNS-2 enhancer and inhibits c-Maf-induced IL-21 production in CD4<sup>+</sup> T cells.

differentiation of Th17 cells while RORyt-deficient mice lack Th17 cell differentiation [21], RORyt is regarded a master regulator of Th17 cells [22]. Additionally, IL-6- and/or IL-21-mediated activation of Stat3 plays a central role in RORyt-induced development of Th17 cells [23-26]. Stat3 binds to intron 1 of *Rorc* gene and promotes trimethylation of histone H3 lysine 4 on *Rorc*, though Stat3 does not activate *Rorc* promoter [27,28]. Regarding the downstream pathways of Stat3 signaling, a number of genes including *Rora*, *Maf*, *Ahr*, *Irf4*, *Batf*, *Nfkbiz*, and *HIF-1* $\alpha$  are activated by Stat3 and involved in Th17 cell differentiation [27,29-34]. Among these transcription factors, HIF-1 activates *Rorc* promoter [34].

Sox5 belongs to the Sox (SRY-related high-mobilitygroup[HMG]-box) family of transcription factors. Sox5 is a member of SoxD family that is composed of Sox6, Sox13, and Sox5[35,36]. SoxD protein has an HMG domain, which mediates the binding to DNA, and two coiled-coil domains, where the first coiled-coil domain induces dimerization of SoxD proteins. SoxD proteins do not have transrepression or transactivation domains and their activity is probably influenced by the molecules they interact with. Sox5-deficient mice die after birth due to a small thoracic cage and cleft secondary palate; this is consistent with the finding that Sox5 is strongly expressed in chondrocytes, neurons, spermatids, and oligodendrocytes. Regarding the relationship between autoimmune diseases and Sox5, it has been recently reported that Sox5 is one of the most significantly upregulated genes in the blood of patients with multiple sclerosis[37]. In addition, DNA microarray analysis showed that Sox5 gene is the most strongly expressed transcription factor in CD4<sup>+</sup> T cells upon IL-6 stimulation. Particularly a novel isoform of Sox5, Sox5t, is expressed in Th17 cells and IL-21producing CD4<sup>+</sup> T cells[38]. Upon stimulation with IL-6, Stat3 induces the expression of c-Maf and Sox5t. Subsequently, c-Maf along with Sox5t induces the expression of RORyt by directly activating the RORyt promoter, suggesting that Sox5t functions downstream of IL-6-Stat3 signaling and upstream of RORyt expression during the differentiation of Th17 cells

(Figure 1).

Additionally, Sox5 plays a crucial role in the onset of Th17 cell-mediated autoimmune models. Experimental autoimmune encephalomyelitis (EAE) is a mouse model of multiple sclerosis mainly caused by Th17 cellmediated autoimmune responses. In this model, CD4<sup>cre</sup> Sox5<sup>fl/fl</sup> mice exhibit significantly reduced inflammatory cell infiltration and clinical scores as compared to the control Sox5<sup>fl/fl</sup> mice. Further, the number of Th17 cells in the brain and spinal cord decreases in CD4<sup>cre</sup> Sox5<sup>fl/fl</sup> mice. In addition, the severity of delayed-type hypersensitivity (DTH) reduces in CD4<sup>cre</sup> Sox5<sup>fl/fl</sup> mice as compared with that in the control Sox5<sup>fl/fl</sup> mice. Furthermore, mice injected with c-Maf- and Sox5texpressing CD4<sup>+</sup> T cells exhibit strong DTH response and IL-17 production in the draining LNs. Collectively, Sox5 is crucially involved in the development of Th17 cell-mediated inflammatory responses.

## **W.** Transcriptional regulation of peripherally induced Treg cells

Treg cells are essential for maintaining immune tolerance in the gut where food antigens and microbiota are present. Among Treg cells, thymus-derived Treg (tTreg) cells are indispensable for maintaining immune tolerance to self-antigens and peripherally induced Treg (pTreg) cells play essential roles in the composition of commensal microbiota and repression of allergic inflammation in mucosa[39]. pTreg cells arise from expression of Foxp3 during CD4<sup>+</sup> T cell differentiation in the periphery and consist of a majority of gut Treg population[40]. In autoimmune colitis models, pTreg cells together with tTreg cells act to restore immune tolerance[41]. These findings suggest that pTreg cells are crucial in suppressing gut inflammation.

Regarding the underlying mechanism of Foxp3 induction during T cell differentiation in the periphery, strong T cell receptor (TCR) stimulation with suboptimal co-stimulation, IL-2, TGF- $\beta$ , microbial metabolites, and retinoic acid induce pTreg cell differentiation both *in vitro* and *in vivo*[42-45].

Under inflammatory conditions, Treg cells enhance

suppressive activity, acquire an activated phenotype, and increase their population [46,47]. Regarding the underlying mechanism of maintaining activated Treg cells, Arvey et al. have shown that Foxp3 modulates the expression of its target genes by inducing repressive histone H3 marks under inflammatory conditions [48]. This study has uncovered the importance of Foxp3mediated transcriptional regulation in activated Treg cells under inflammatory conditions.

RNA sequencing analysis of Treg cells in dextran sulfate sodium-induced colitic mice reveals that Sex determining region Y box 12 (Sox12) is the only transcription factor that is significantly induced in Treg cells[49]. Sox12 is a member of the SoxC family that is composed of Sox4, Sox11, and Sox12. SoxC proteins have a C-terminal transactivation domain and an N-terminal HMG box domain that mediates binding to DNA. These proteins play crucial roles in the development of the nerve system, pancreas, and kidneys [50]. Regarding immunological aspects, Sox4 is involved in survival of the B cell precursor and differentiation of Th2 cells[51]. Although Sox12 and Sox4 but not Sox11 are expressed in Treg cells, TCR stimulation strongly upregulates Sox12 expression but downregulates Sox4 expression in Treg cells. Upon TCR stimulation, NFAT is activated and it binds upstream of exon 1 of Sox12 gene locus. Thus, TCR-NFAT signaling is important for inducing Sox12 expression in CD4<sup>+</sup> T cells[49]. In addition, Sox12 is involved in the differentiation of pTreg cells under inflammatory conditions in colitic mice, though Sox12 is not essential for the development of tTreg cells. Moreover, overexpression of Sox12 is sufficient to induce Foxp3 expression in CD4<sup>+</sup> T cells even in the absence of IL-2 or TGF- $\beta$  where Sox12 binds to the Foxp3 promoter and drives its transcription (Figure 1). In summary, TCR-NFAT signaling induces the development of pTreg cells in colitic mice partly through Sox12 induction.

## Concluding remarks and future directions

Recent studies have revealed how Sox family proteins regulate the development of Th17 cells, Treg

cells, as well as the onset of autoimmune diseases. Sox5, a protein of SoxD family, induces Th17 cell differentiation. Sox12, a protein of SoxC family, is required for pTreg differentiation. However, the role of Sox4, another protein of the SoxC family in the regulation of Treg cells is still largely unknown. Thus, further research on elucidating the mechanism underlying development of Treg cells is needed.

## **Financial support**

This work was supported in part by Grants-in-Aid for Scientific Research from the Ministry of Education, Culture, Sports, Science and Technology, the Japanese Government, and LGS (Leading Graduate School at Chiba University) Program, MEXT, and Institute for Global Prominent Research, Chiba University, Japan.

## **Conflict of interest**

The author declares no conflict of interest associated with this manuscript.

### Ethical approval

Not applicable.

#### Data availability

Not applicable.

### Acknowledgements

The author expresses gratitude to Prof. Hiroshi Nakajima for critically reading the manuscript.

#### References

- Spolski R. and Leonard WJ. (2008) Interleukin-21: basic biology and implications for cancer and autoimmunity. Annu Rev Immunol 26, 57-79.
- 2) Sutherland AP, Van Belle T, Wurster AL, Suto A, Michaud M, Zhang D, Grusby M J, von Herrath M. (2009) Interleukin-21 is required for the development of

type 1 diabetes in NOD mice. Diabetes 58, 1144-55.

- 3) King C, Ilic A, Koelsch K, Sarvetnick N. (2004) Homeostatic expansion of T cells during immune insufficiency generates autoimmunity. Cell 117, 265-77.
- 4) Vinuesa CG, Cook MC, Angelucci C, Athanasopoulos V, Rui L, Hill KM, Yu D, Domaschenz H, Whittle B, Lambe T, Roberts IS, Copley RR, Bell JI, Cornall RJ, Goodnow CC. (2005) A RING-type ubiquitin ligase family member required to repress follicular helper T cells and autoimmunity. Nature 435, 452-8.
- 5) Young DA, Hegen M, Ma HL, Whitters MJ, Albert LM, Lowe L, Senices M, Wu PW, Sibley B, Leathurby Y, Brown TP, Nickerson-Nutter C, Keith JC, Collins M. (2007) Blockade of the interleukin-21/interleukin-21 receptor pathway ameliorates disease in animal models of rheumatoid arthritis. Arthritis Rheum 56, 1152-63.
- 6) Herber D, Brown TP, Liang S, Young DA, Collins M, Dunussi-Joannopoulos K. (2007) IL-21 has a pathogenic role in a lupus-prone mouse model and its blockade with IL-21R.Fc reduces disease progression. J Immunol 178, 3822-30.
- 7) Wildin RS, Smyk-Pearson S, Filipovich AH. (2002) Clinical and molecular features of the immunodysregulation, polyendocrinopathy, enteropathy, X linked (IPEX) syndrome. J Med Genet 39, 537-45.
- 8) Fontenot JD, Gavin MA, Rudensky AY. (2003) Foxp3 programs the development and function of CD4<sup>+</sup>CD25<sup>+</sup> regulatory T cells. Nat Immunol 4, 330-6.
- 9) Blair PJ, Bultman SJ, Haas JC, Rouse BT, Wilkinson JE, Godfrey VL. (1994) CD4<sup>+</sup>CD8<sup>-</sup> T cells are the effector cells in disease pathogenesis in the scurfy (sf) mouse. J Immunol 153, 3764-74.
- 10) Iwamoto T, Suto A, Tanaka S, Takatori H, Suzuki K, Iwamoto I. Nakajima H. (2014) Interleukin-21-producing c-Maf-expressing CD4<sup>+</sup> T cells induce effector CD8<sup>+</sup> T cells and enhance autoimmune inflammation in scurfy mice. Arthritis Rheumatol 66, 2079-90.
- Sharma R, Jarjour WN, Zheng L, Gaskin F, Fu SM, Ju ST. (2007) Large functional repertoire of regulatory T-cell suppressible autoimmune T cells in scurfy mice. J Autoimmun 29, 10-9.
- 12) Young NA, Sharma R, Friedman AK, Kaffenberger BH, Bolon B, Jarjour WN. (2013) Aberrant muscle antigen exposure in mice is sufficient to cause myositis in a Treg cell-deficient milieu. Arthritis Rheum 65, 3259-70.
- Simon JP, Marie I, Jouen F, Boyer O, Martinet J. (2016) Autoimmune Myopathies: Where Do We Stand? Front Immunol 7, 234.
- Morita R, Schmitt N, Bentebibel SE, Ranganathan R, Bourdery L, Zurawski G, Foucat E, Dullaers M, Oh S, Sabzghabaei N, Lavecchio EM, Punaro M, Pascual V, Banchereau J, Ueno H. (2011) Human blood CXCR5 (+) CD4 (+) T cells are counterparts of T follicular cells and contain specific subsets that differentially support antibody secretion. Immunity 34, 108-21.
- 15) Kageyama T, Suto A, Iwamoto T, Tanaka S, Suehiro K, Yokoyama Y, Saku A, Furuta S, Ikeda K, Suzuki

K, Hirose K, Nakajima H. (2017) IL-21 Exacerbates Autoimmune Myositis by Enhancing the Accumulation of GM-CSF–Producing  $\gamma\delta T$  Cells in the Muscle. ImmunoHorizons 1, 176-87.

- 16) Suto A, Kashiwakuma D, Kagami S, Hirose K, Watanabe N, Yokote K, Saito Y, Nakayama T, Grusby MJ, Iwamoto I, Nakajima H. (2008) Development and characterization of IL-21-producing CD4<sup>+</sup> T cells. J Exp Med 205, 1369-79.
- Mehta DS, Wurster AL, Grusby MJ. (2004) Biology of IL-21 and the IL-21 receptor. Immunol Rev 202, 84-95.
- 18) Chen Q, Yang W, Gupta S, Biswas P, Smith P, Bhagat G, Pernis AB. (2008) IRF-4-binding protein inhibits interleukin-17 and interleukin-21 production by controlling the activity of IRF-4 transcription factor. Immunity 29, 899-911.
- 19) Nurieva R, Yang XO, Martinez G, Zhang Y, Panopoulos AD, Ma L, Schluns K, Tian Q, Watowich SS, Jetten AM, Dong C. (2007) Essential autocrine regulation by IL-21 in the generation of inflammatory T cells. Nature 448, 480-3.
- 20) Hiramatsu Y, Suto A, Kashiwakuma D, Kanari H, Kagami S, Ikeda K, Hirose K, Watanabe N, Grusby MJ, Iwamoto I, Nakajima H. (2010) c-Maf activates the promoter and enhancer of the IL-21 gene, and TGFbeta inhibits c-Maf-induced IL-21 production in CD4<sup>+</sup> T cells. J Leukoc Biol 87, 703-12.
- 21) Ivanov II, McKenzie BS, Zhou L, Tadokoro CE, Lepelley A, Lafaille JJ, Cua DJ, Littman DR. (2006) The orphan nuclear receptor RORgammat directs the differentiation program of proinflammatory IL-17<sup>+</sup> T helper cells. Cell 126, 1121-33.
- 22) Ciofani M, Madar A, Galan C, Sellars M, Mace K, Pauli F, Agarwal A, Huang W, Parkurst CN, Muratet M, Newberry KM, Meadows S, Greenfield A, Yang Y, Jain, P, Kirigin FK, Birchmeier C, Wagner EF, Murphy KM, Myers RM, Bonneau R, Littman DR. (2012) A validated regulatory network for Th17 cell specification. Cell 151, 289-303.
- 23) Zhou L, Ivanov II, Spolski R, Min R, Shenderov K, Egawa T, Levy DE, Leonard WJ, Littman DR. (2007) IL-6 programs T (H)-17 cell differentiation by promoting sequential engagement of the IL-21 and IL-23 pathways. Nat Immunol 8, 967-74.
- 24) Mathur AN, Chang HC, Zisoulis DG, Stritesky GL, Yu Q, O'Malley JT, Kapur R, Levy DE, Kansas GS, Kaplan MH. (2007) Stat3 and Stat4 direct development of IL-17-secreting Th cells. J Immunol 178, 4901-7.
- 25) Laurence A, Tato CM, Davidson TS, Kanno Y, Chen Z, Yao Z, Blank RB, Meylan F, Siegel R, Hennighausen L, Shevach EM, O'shea JJ. (2007) Interleukin-2 signaling via STAT5 constrains T helper 17 cell generation. Immunity 26, 371-81.
- 26) Yang XO, Panopoulos AD, Nurieva R, Chang SH, Wang D, Watowich SS, Dong C. (2007) STAT3 regulates cytokine-mediated generation of inflammatory helper T cells. J Biol Chem 282, 9358-63.

22

- 27) Durant L, Watford WT, Ramos HL, Laurence A, Vahedi G, Wei L, Takahashi H, Sun HW, Kanno Y, Powrie F, O'Shea JJ. (2010) Diverse targets of the transcription factor STAT3 contribute to T cell pathogenicity and homeostasis. Immunity 32, 605-15.
- 28) Lazarevic V, Chen X, Shim JH, Hwang ES, Jang E, Bolm AN, Oukka M, Kuchroo VK, Glimcher LH. (2011) T-bet represses T (H) 17 differentiation by preventing Runx1-mediated activation of the gene encoding RORγt. Nat Immunol 12, 96-104.
- 29) Yang XO, Pappu BP, Nurieva R, Akimzhanov A, Kang HS, Chung Y, Ma L, Shah B, Panopoulos AD, Schluns KS, Watowich SS, Tian Q, Jetten AM, Dong C. (2008) T helper 17 lineage differentiation is programmed by orphan nuclear receptors ROR alpha and ROR gamma. Immunity 28, 29-39.
- 30) Schraml BU, Hildner K, Ise W, Lee WL, Smith WA, Solomon B, Sahota G, Sim J, Mukasa R, Cemerski S, Hatton RD, Stormo GD, Weaver CT, Russell JH, Murphy TL, Murphy KM. (2009) The AP-1 transcription factor Batf controls T (H) 17 differentiation. Nature 460, 405-9.
- 31) Brüstle A, Heink S, Huber M, Rosenplänter C, Stadelmann C, Yu P, Arpaia E, Mak TW, Kamradt T, Lohoff M. (2007) The development of inflammatory T (H)-17 cells requires interferon-regulatory factor 4. Nat Immunol 8, 958-66.
- 32) Veldhoen M, Hirota K, Westendorf AM, Buer J, Dumoutier L, Renauld JC, Stockinger B. (2008) The aryl hydrocarbon receptor links TH17-cell-mediated autoimmunity to environmental toxins. Nature 453, 106-9.
- 33) Bauquet AT, Jin H, Paterson AM, Mitsdoerffer M, Ho IC, Sharpe AH, Kuchroo VK. (2009) The costimulatory molecule ICOS regulates the expression of c-Maf and IL-21 in the development of follicular T helper cells and TH-17 cells. Nat Immunol 10, 167-75.
- 34) Dang EV, Barbi J, Yang HY, Jinasena D, Yu H, Zheng Y, Bordman Z, Fu J, Kim Y, Yen HR, Luo W, Zeller K, Shimoda L, Topalian SL, Semenza GL, Dang CV, Pardoll DM, Pan F. (2011) Control of T (H) 17/T (reg) balance by hypoxia-inducible factor 1. Cell 146, 772-84.
- 35) Lefebvre V, Li P, de Crombrugghe B. (1998) A new long form of Sox5 (L-Sox5), Sox6 and Sox9 are coexpressed in chondrogenesis and cooperatively activate the type II collagen gene. EMBO J 17, 5718-33.
- 36) Lefebvre V. (2010) The SoxD transcription factors--Sox5, Sox6, and Sox13--are key cell fate modulators. Int J Biochem Cell Biol 42, 429-32.
- 37) Riveros C, Mellor D, Gandhi KS, McKay FC, Cox MB, Berretta R, Vaezpour SY, Inostroza-Ponta M, Broadley SA, Heard RN, Vucic S, Stewart GJ, Williams DW, Scott RJ, Lechner-Scott J, Booth DR, Moscato P, ANZgene Multiple Sclerosis Genetics Consortium. (2010) A transcription factor map as revealed by a genomewide gene expression analysis of whole-blood mRNA transcriptome in multiple sclerosis. PLoS One 5, e14176.

- 38) Tanaka S, Suto A, Iwamoto T, Kashiwakuma D, Kagami S, Suzuki, K, Takatori H, Tamachi T, Hirose K, Onodera A, Suzuki J, Ohara O, Yamashita M, Nakayama T, Nakajima H. (2014) Sox5 and c-Maf cooperatively induce Th17 cell differentiation via RORgammat induction as downstream targets of Stat3. J Exp Med 211, 1857-74.
- 39) Josefowicz SZ, Niec RE, Kim HY, Treuting P, Chinen T, Zheng Y, Umetsu DT, Rudensky AY. (2012) Extrathymically generated regulatory T cells control mucosal TH2 inflammation. Nature 482, 395-9.
- Ai TL, Solomon BD, Hsieh CS. (2014) T-cell selection and intestinal homeostasis. Immunological reviews 259, 60-74.
- 41) Haribhai D, Lin W, Edwards B, Ziegelbauer J, Salzman NH, Carlson MR, Li SH, Simpson PM, Chatila TA, Williams CB. (2009) A central role for induced regulatory T cells in tolerance induction in experimental colitis. Journal of immunology 182, 3461-8.
- 42) Bilate AM and Lafaille JJ. (2012) Induced CD4<sup>+</sup> Foxp3<sup>+</sup> regulatory T cells in immune tolerance. Annu Rev Immunol 30, 733-58.
- 43) Furusawa Y, Obata Y, Fukuda S, Endo TA, Nakato G, Takahashi D, Nakanishi Y, Uetake C, Kato K, Kato T, Takahashi M, Fukuda NN, Murakami S, Miyauchi E, Hino S, Atarashi K, Onawa S, Fujimura Y, Lockett T, Clarke JM, Topping DL, Tomita M, Hori S, Ohara O, Morita T, Koseki H, Kikuchi J, Honda K, Hase K, Ohno H. (2013) Commensal microbe-derived butyrate induces the differentiation of colonic regulatory T cells. Nature 504, 446-50.
- 44) Arpaia N, Campbell C, Fan X, Dikiy S, van der Veeken J, deRoos P, Liu H, Cross JR, Pfeffer K, Coffer PJ, Rudensky AY. (2013) Metabolites produced by commensal bacteria promote peripheral regulatory T-cell generation. Nature 504, 451-5.
- 45) Josefowicz SZ, Lu LF, Rudensky AY. (2012) Regulatory T cells: mechanisms of differentiation and function. Annu Rev Immunol 30, 531-64.
- 46) Shafiani S, Dinh C, Ertelt JM, Moguche AO, Siddiqui I, Smigiel KS, Sharma P, Campbell DJ, Way SS, Urdahl KB. (2013) Pathogen-specific Treg cells expand early during mycobacterium tuberculosis infection but are later eliminated in response to Interleukin-12. Immunity 38, 1261-70.
- 47) Rosenblum MD, Gratz IK, Paw JS, Lee K, Marshak-Rothstein A, Abbas AK. (2011) Response to self antigen imprints regulatory memory in tissues. Nature 480, 538-42.
- 48) Arvey A, van der Veeken J, Samstein RM, Feng Y, Stamatoyannopoulos JA, Rudensky AY. (2014) Inflammation-induced repression of chromatin bound by the transcription factor Foxp3 in regulatory T cells. Nature immunology 15, 580-7.
- 49) Tanaka S, Suto A, Iwamoto T, Kageyama T, Tamachi T, Takatori H, Suzuki K, Hirose K, Ohara O, Lefebvre V, Nakajima H. (2018) Sox12 promotes T reg

differentiation in the periphery during colitis. J Exp Med 215, 2509-19.

- 50) Penzo-Mendez AI. (2010) Critical roles for SoxC transcription factors in development and cancer. Int J Biochem Cell Biol 42, 425-8.
- 51) Kuwahara M, Yamashita M, Shinoda K, Tofukuji S,

Onodera A. Shinnakasu R, Motohashi S, Hosokawa H, Tumes D, Iwamura C, Lefebvre V, Nakayama T. (2012) The transcription factor Sox4 is a downstream target of signaling by the cytokine TGF- $\beta$  and suppresses T (H) 2 differentiation. Nat Immunol 13, 778-86.